



REVIEW

Open Access

# Macrophages and angiogenesis: a role for Wnt signaling

Andrew C Newman<sup>1</sup> and Christopher C W Hughes<sup>1,2,3\*</sup>

## Abstract

Macrophages regulate many developmental and pathological processes in both embryonic and adult tissues, and recent studies have shown a significant role in angiogenesis. Similarly, Wnt signaling is fundamental to tissue morphogenesis and also has a role in vascular development. In this review, we summarize recent advances in the field of macrophage-regulated angiogenesis, with a focus on the role of macrophage-derived Wnt ligands. We review data that provide both direct and indirect evidence for macrophage-derived Wnt regulation of physiologic and pathologic angiogenesis. Finally, we propose that Wnt signaling plays a central role in differentiation of tumor associated and wound infiltrating macrophages to a proangiogenic phenotype.

**Keywords:** Macrophage, Angiogenesis, Wnt

## Introduction

Angiogenesis is the growth of new blood vessels from the existing vasculature. This process plays critical roles in both development and pathological conditions such as tumor growth and diabetic retinopathy [1]. During angiogenesis, endothelial cells (EC) coordinate signals derived from a multitude of stromal cells in close proximity to the blood vessels, including fibroblasts, smooth muscle cells/pericytes and macrophages [2]. Growth factors, proteases and extracellular matrix (ECM) components derived from these stromal cells participate in the process [1]. In this review, we will focus on the role of macrophage-derived Wnt signals in regulating angiogenesis.

Members of the Wnt protein family have been shown to regulate diverse biological processes including cell proliferation, apoptosis, polarity, differentiation and the maintenance of pluripotency in stem cells [3]. Wnt proteins have been studied extensively, but only recently has their function in angiogenesis begun to be elucidated. Macrophages comprise a diverse group of cells from the mononuclear phagocytic lineage [4]. They are highly plastic, exhibiting dramatic changes in phenotype in response to various stimuli and, in addition, are believed to exist in a

multitude of subpopulations [4-6]. This review is not intended to be an overview of different macrophage phenotypes and subpopulations, instead, we highlight studies showing an angiogenic function for a variety of the different subpopulations of macrophages. We will employ a general definition of macrophages as any non-dendritic cell of monoblast origin, which includes monocytes and the specialized tissue-resident macrophages of the central nervous system, microglial cells. Increasing evidence suggests that macrophages mediate their effects on angiogenesis, in part, through secretion of members of the Wnt family of secreted glycoproteins.

## The role of Wnts in angiogenesis

In humans, Wnt signaling is mediated by a family of 19 secreted Wnt glycoproteins and 10 transmembrane Frizzled (Fzd) receptors [7]. Although the distinction is becoming less clear [8,9] Wnt signaling is generally broken down into the canonical, Wnt/ $\beta$ -catenin pathway, and the non-canonical, Wnt/Calcium and planar cell polarity (PCP) pathway. In the canonical Wnt pathway, Wnt ligand binding to Fzd and the co-receptors, LDL-receptor-related proteins-5 or -6 (Lrp5/6), leads to cytosolic  $\beta$ -catenin accumulation.  $\beta$ -catenin then translocates to the nucleus where it binds to transcription factors of the lymphocyte enhancing factor (LEF)/T-cell factor (TCF) family, thereby activating transcription of a number of target genes [7,10]. Non-canonical Wnts

\* Correspondence: cchughes@uci.edu

<sup>1</sup>The Department of Molecular Biology and Biochemistry, University of California Irvine, Irvine, CA 92697, USA

<sup>2</sup>The Department of Biomedical Engineering, University of California Irvine, Irvine, CA 92697, USA

Full list of author information is available at the end of the article

signal through Fzd receptors as well as members of the receptor tyrosine kinase-like orphan receptor (Ror) family and the Wnt modifier, receptor-like tyrosine kinase (Ryk). This pathway leads to changes in cell polarity and migration and is mediated by  $Ca^{2+}$  influx as well as activation of the small GTPases, RhoA, Cdc42 and Rac [11,12]. A detailed overview of Wnt signaling is beyond the scope of this review and the reader is referred to many excellent reviews [7,8,10,11] as well as the Wnt Gene Homepage at <http://www.stanford.edu/~rnusse/wntwindow.html>.

Evidence for Wnt signals regulating angiogenesis is provided firstly by reports of ECs expressing a number of Wnt receptors including Fzd1, Fzd2, Fzd4, Fzd5, Fzd6, Fzd7, Fzd9, Fzd10, Lrp5, Lrp6 and the Wnt signaling modulator, Ryk [13-16]. It is not surprising then, that there are numerous reports showing that EC respond to Wnt proteins in vitro. Wnt1, Wnt3a and Wnt5a have all been shown to control EC proliferation and in some cases migration [17-23], two processes critical for angiogenesis. In addition, the Wnt antagonist, secreted frizzled-related protein 1 (sFRP1) reduces EC proliferation [24] and canonical Wnt signaling was shown to promote tube-like formation on Matrigel [25]. Masckauchan et al. have shown that Wnt5a induces expression of Tie-2 in human umbilical vein endothelial cells (HUVEC), a receptor tyrosine kinase involved in EC survival and maturation [19] and we have independently confirmed this finding in our lab (unpublished observations). Thus, it is clear that Wnt signaling pathways can regulate angiogenesis in vitro.

The story is much the same in vivo, with several reports of active Wnt signaling in EC undergoing angiogenesis [26-33]. Moreover, many Wnt signaling pathway alterations lead to vascular defects.  $\beta$ -catenin nuclear localization has been observed in capillaries during human development [28] and various reporter mice have shown that canonical Wnt signaling is active in EC during developmental angiogenesis [27,30,31].  $\beta$ -catenin has also been shown to accumulate in the nucleus of EC undergoing pathological angiogenesis in rat models of myocardial infarction [26] and glioma [32] as well as in human tumors of the central nervous system [29,33]. Of note,  $\beta$ -catenin nuclear localization is rarely seen in adult quiescent EC [26,29,32,33]. Changes in Wnt signaling pathways have also been correlated with angiogenesis. For example, endothelial-specific  $\beta$ -catenin gain-of-function mutant mice are embryonic lethal and display decreased vessel lengths and increased lumen diameter [34]. These mice also display upregulated Notch signaling, another important mediator of vessel growth [34]. Injection of cells overexpressing Wnt1 and Wnt3a into the chick paraxial mesoderm leads to increased vascular density [35], and disruption of the Wnt2 gene in mice

results in placental vascular defects with a decrease in the number of fetal capillaries [36].

Disturbing Wnt receptor expression in vivo can also lead to vascular defects, with the most well-studied receptors being Fzd4 and Fzd5. Fzd4 loss-of-function mutations have been linked to familial exudative vitreoretinopathy (FEVR), a disease characterized by abnormal retinal vasculature [37]. Furthermore, Fzd4 deletion in mice has been shown to alter vessel formation in the cerebellum and the retina [38]. However, at least in the retina, one Fzd4 ligand is Norrin, which is not a Wnt family member, but does activate the canonical Wnt signaling pathway [38]. In a recent study, Descamps et al. showed that Fzd4<sup>-/-</sup> mice display significant reductions in arteriole density in the heart and kidney as well as impaired angiogenesis in a mouse model of hind limb ischemia [39]. The evidence presented in that study suggests that the Fzd4 ligand is a non-canonical Wnt. Fzd5 knockout (KO) mice are embryonic lethal due to defects in yolk sac angiogenesis. Specifically, EC proliferation is impaired and an incomplete capillary plexus is formed [40]. A more recent study using a conditional genetic deletion of Fzd5 to bypass the perinatal lethality observed in the original KO mouse, shows that this Wnt receptor also regulates blood vessel development [41].

Clearly there is a plethora of evidence implicating Wnt signaling in modulating angiogenesis. For a more extensive overview of the literature, the reader is referred to several recent reviews [42-44].

### Macrophages in angiogenesis

There is substantial evidence that macrophages play a significant role in both physiological and pathological angiogenesis [4,45,46]. In mouse development the first macrophages appear at 7.5 days post-coitum (dpc) and starting between 8 and 9.5 dpc they can be found in both extra-embryonic and embryonic tissue [47], placing them in prime position to regulate angiogenesis. In support of this idea, embryonic macrophages, compared to circulating adult macrophages, have been shown to have "wound healing" and "angiogenesis" gene expression signatures [48]. Macrophages colonize the mouse embryonic hind-brain independently of blood vessel formation and Fantin et al. showed that these tissue-resident macrophages associated with angiogenic tip cells during the same time period that neighboring vessels were anastomosing with one another [49]. Ablation of macrophages, using both PU.1<sup>-/-</sup> and Csf-1<sup>op/op</sup> mice, resulted in a decreased number of vessel intersections in the hindbrain, providing the first evidence that macrophages actively participate in vessel anastomosis [49]. Interestingly, these tissue-resident macrophages were shown to express Tie-2 and NRP1, two genes that are known to be significantly upregulated in proangiogenic macrophages [47]. Also of note, the effect of these macrophages on angiogenesis is not mediated by

vascular endothelial growth factor-A (VEGF-A) secretion [49]. However, the study does not rule out the possibility that macrophages secrete other VEGF isoforms that contribute to angiogenesis. Evidence for macrophages regulating vessel anastomosis in other tissues comes from studies in the mouse retina [49-51]. Outtz and colleagues provided evidence that macrophage Notch1 regulates this process [50] and studies from Rymo et al. indicate that microglial-secreted factors other than VEGF-A increase vessel sprouting and branching in the rat aortic ring model of angiogenesis [51].

Macrophages, given their ability to migrate within virtually all tissues of the body, are also an ideal cell to regulate angiogenesis during tissue injury and repair. Using a transgenic mouse model of ischemic cardiomyopathy, Moldovan et al. showed that macrophages carved out tunnels in the ECM, thereby providing avenues for subsequent capillary infiltration [52]. Macrophages are actively recruited to wound sites and macrophage-depletion during early phases of the wound healing process leads to reduced formation of vascularized granulation tissue, whereas depletion during later phases caused severe hemorrhage and prevented wound closure [53]. These findings implicate macrophages in mediating both the initial and maturation stages of angiogenesis. A separate study showed that wound healing was delayed in macrophage-deficient *Csf-1<sup>op/op</sup>* mice and this correlated with decreased vascular density [54]. Thus, although it is well known that macrophages serve an immune function during tissue repair, it is also clear that they play a trophic role through the regulation of angiogenesis.

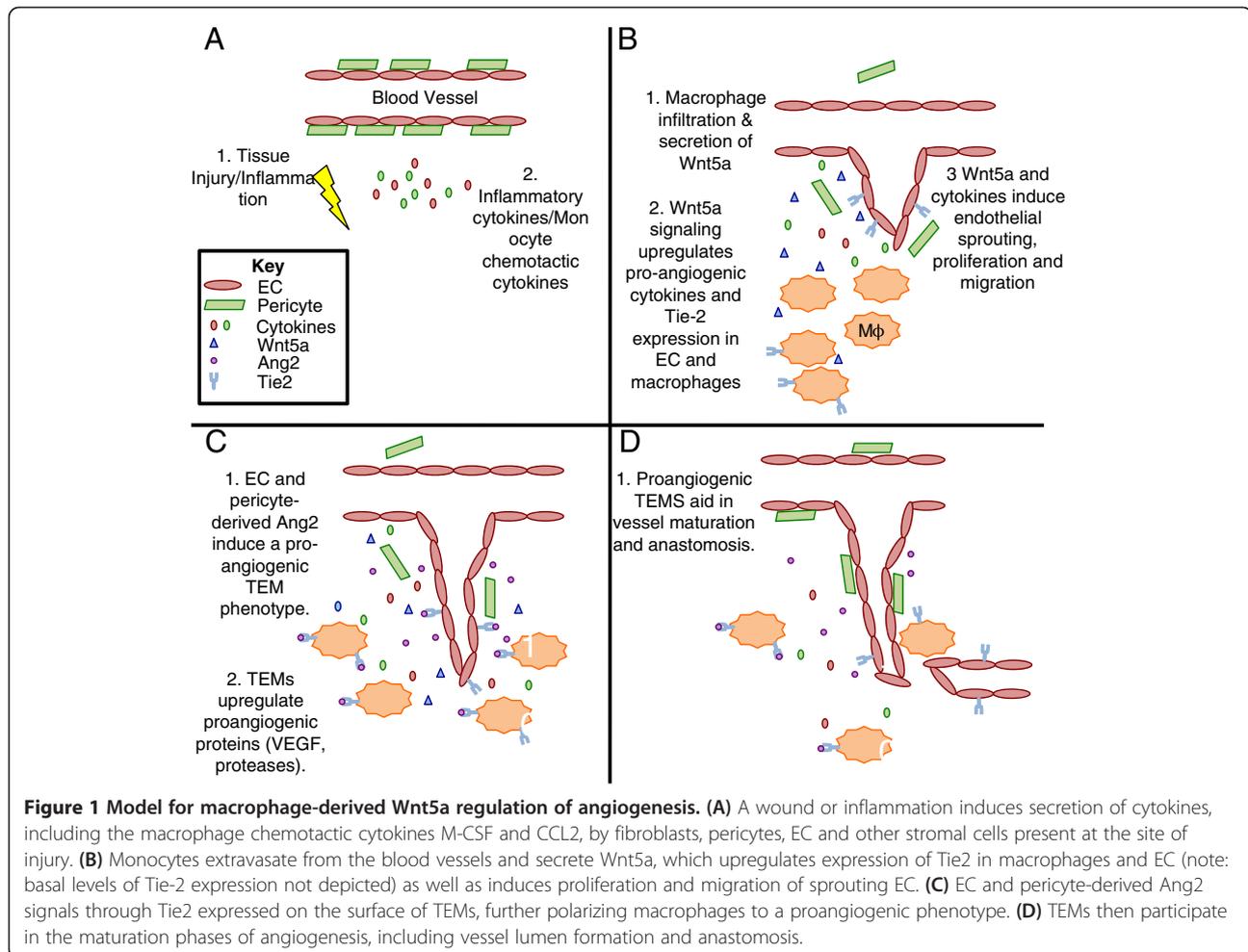
A rapidly developing field is the study of tumor-associated macrophages (TAMs) and their role in promoting tumor progression. It comes as no surprise that TAMs mediate some of their effects on tumor growth by regulating angiogenesis [45,46,55,56]. TAMs can be recruited to tumors by a number of different cytokines, including CCL2 and CSF-1/M-CSF [57-59]. TAMs have been shown to regulate tumor angiogenesis in a number of different tissues including bone, brain, breast, cervix, colon and lung, among others [60-66]. They have been shown to mediate angiogenesis in part by secretion of VEGF, along with proteases, such as matrix metalloproteinases (MMPs) [64,67]. Interestingly, studies by De Palma and colleagues have shown that the majority of proangiogenic TAMs in tumors are Tie-2 expressing monocytes (TEMs) [47,68], implicating Tie-2 as an important marker of proangiogenic macrophages. Indeed, Angiopoietin-2 (Ang-2) signaling via Tie-2 in TEMs has been shown to upregulate proangiogenic genes [69]. Although TEMs have been shown to exist as a subset of circulating monocytes [68], studies have shown that conditions of hypoxia can upregulate Tie-2 expression in monocytes [70]. As in EC [19], Tie-2 expression in

macrophages may also be regulated by Wnt5a. It is clear that macrophages regulate angiogenesis through multiple mechanisms, however, a detailed discussion on the role of macrophage-derived cytokines in angiogenesis is beyond the scope of this review.

### **Macrophage-derived Wnts regulate angiogenesis**

Based on the studies outlined above, it is evident that both macrophages and secreted Wnt proteins regulate angiogenesis. It may, therefore, be reasonable to hypothesize that Wnt ligands mediate some of the effects that macrophages have on angiogenesis, and indeed, macrophages do express Wnt ligands. Interferon- $\gamma$  (IFN- $\gamma$ ) and lipopolysaccharide (LPS), two potent inducers of inflammation, lead to significant upregulation of Wnt5a transcript and protein levels in macrophages [71]. Furthermore, Wnt5a can signal in an autocrine manner to induce expression of several pro-inflammatory cytokines in macrophages including IL-6, IL-8 and IL-1 $\beta$  [71], all of which have been shown to be pro-angiogenic [72-75]. Given that Wnt5a has also been shown to induce expression of several inflammatory cytokines in EC, including IL-6 and IL-8 [76] combined with the direct effects that Wnt5a has on EC proliferation, migration and Tie-2 expression [18,19,22], it is reasonable to hypothesize that macrophage-derived Wnt5a regulates angiogenesis via both direct and indirect mechanisms and at multiple levels. Moreover, as Wnt5a has also been shown to upregulate expression of the macrophage chemotactic protein, CCL2, in EC [76], it is possible that macrophage-derived Wnt5a could indirectly drive additional rounds of macrophage recruitment.

Aside from inflammatory angiogenesis, research in cancer biology also points to a role for macrophage-derived Wnts in regulating tumor angiogenesis. Smith et al. looked at Wnt expression in 14 matched cases of normal, adenomatous and malignant colorectal tissues and reported upregulation of Wnt gene expression in TAMs, in particular, Wnt2 and Wnt5a, during the progression from normal through adenoma to carcinoma in colorectal tissue [77]. Of note, the angiogenic switch in colorectal cancer is thought to occur during the progression from adenoma to carcinoma [78]. As mentioned earlier, TAMs play an essential role in breast cancer angiogenesis [60,61]. Ojalvo et al. separated "invasive" TAMs from general TAMs using an *in vivo* migration assay [79] combined with fluorescent activated cell sorting (FACS) [80], and used gene expression analysis to show that "invasive" TAMs were enriched for Wnt signaling molecules [80]. Based on these findings and others, Qian and Pollard recently hypothesized that this subset of Wnt-expressing-TAMs may link angiogenesis and tumor invasion [46]. Further investigation is needed to confirm the role of macrophage-derived Wnts in regulating colorectal and breast tumor angiogenesis.



Direct evidence for the involvement of macrophage-derived Wnts in regulating hyaloid vascular remodeling and developmental angiogenesis in the retina comes from studies by Lang and colleagues. The hyaloid vessel system is laid down during development of the eye and requires remodeling after birth to allow for unobstructed vision [81]. Macrophages are in close contact with these vessels and genetic approaches that deplete macrophages lead to a loss of remodeling and the persistence of these vessels post-natally [82]. Further examination revealed that macrophage secretion of the canonical Wnt protein, Wnt7b, is required for this process [83]. Specifically, pericyte secretion of Ang-2 induces Wnt7b expression in macrophages; Wnt7b then stimulates vascular EC entry into the S phase of the cell cycle. Pericyte secretion of Ang-2 inhibits Ang-1-mediated survival signaling in EC and also signals through  $\beta$ -catenin to induce cell cycle entry and subsequent apoptosis [83,84]. The authors hypothesize that the need for macrophage-derived Wnt7b allows for coordination of apoptosis and phagocytosis [84].

Non-canonical Wnts secreted by macrophages also regulate vessel branching in the mouse retina. Shortly

after birth, vessels from the superficial vascular plexus on the surface of the retina sprout vertically, down through the ganglion. Upon reaching the outer edge of the inner nuclear layer, EC turn and branch to form the deep vascular plexus [85]. Stefater et al. showed that retinal myeloid cells (RMC) are in close contact with tip cells of EC sprouts at the point of branching and remained associated after EC extend within the plane of the deep retinal layer [86]. They further showed that this population of RMCs is different than RMCs found in the superficial vascular layer. Specifically, they express the non-canonical Wnt ligands, Wnt5a and Wnt11, whereas superficial RMCs do not [86]. The authors also demonstrated that myeloid-specific deletion of the gene for the Wnt ligand transporter Wls, a protein required for Wnt secretion [87], resulted in increased vascular density of the deep vascular plexus, a phenotype shared by Wnt5a<sup>+/-</sup> and Wnt11<sup>+/-</sup> heterozygotes. The mechanism was shown to involve an autocrine-signaling axis where macrophage-derived Wnt5a and Wnt11 induce secretion of the soluble VEGF inhibitor Flt1 [88,89] that, in turn, inhibits vessel branching [86].

It is clear that Wnt signaling is important for angiogenic regulation by macrophages and we anticipate that future research in this field will reveal an even more crucial role for these proteins in this process.

### Conclusions and perspective

We have laid out evidence indicating that macrophages regulate angiogenesis, in part, through secreting and responding to Wnt signaling glycoproteins. This is true in both developmental and pathological angiogenesis. It is interesting and perhaps not unexpected that in some cases Wnts seem to be pro-angiogenic, as is the case for Wnt5a-induced EC proliferation and migration [19,20], and in other cases inhibitory, for example Wnt5a inhibition of vessel branching in the retinal deep vascular plexus [86]. One possible explanation for this relates to the fact that angiogenesis is a *series* of events with different morphological changes required at different stages. Therefore, the same protein may inhibit or induce angiogenesis depending on the stage of angiogenesis at which it is present. We also reason that these differences are due to differential Wnt receptor expression on the various cells responding to the signal as well as different signaling proteins that are present in the microenvironment. Coordinated crosstalk of Wnt signaling pathways with other pathways in EC such as Notch/Dll4, VEGF, Ang1/2 and focal adhesion kinase (FAK) [90] ultimately will determine the nature of the response. Furthermore, the cell population in a given tissue will likely alter the angiogenic response to macrophage-derived Wnts. We have discussed Wnt signaling in ECs and macrophages, but it is likely that Wnt binding to fibroblasts, smooth muscle cells and pericytes alters their phenotype as well. In the cases described above, differences in the effect of Wnt5a on angiogenesis could be explained by which cells bind the Wnt5a ligand. Wnt5a binding directly on the EC induces proliferation and migration, whereas Wnt5a signaling in an autocrine manner in macrophages leads to upregulation of the soluble VEGF inhibitor, Flt1, to reduce vessel branching.

Based on the studies that we have presented in this review, we hypothesize that Wnt5a is a crucial mediator of macrophage phenotype. It is known that Wnt5a induces Tie-2 expression in EC [19] and it would be interesting to determine if macrophages respond in the same manner, especially as TEMs are known to be critical mediators of angiogenesis [47,68,69]. This would implicate Wnt5a as an important determinant of macrophage differentiation from an inflammatory function to a more trophic function. We propose a model where wounding or inflammation leads to a release of inflammatory cytokines, including the macrophage chemotactic proteins CCL2 and M-CSF (Figure 1A). Upon recruitment, macrophages release Wnt5a, which induces EC proliferation and migration, but also upregulates Tie-2 expression in macrophages and EC (Figure 1B).

These Tie-2 expressing macrophages (TEM) respond to Ang-2 secreted by EC and pericytes and polarize into a more proangiogenic macrophage [69] (Figure 1C). During later stages of angiogenesis these TEMs aid in vessel anastomosis and maturation (Figure 1D).

Additionally, studies of TAMs show that “invasive” TAMs in breast cancer are enriched for Wnt signaling pathway components [80] and colon cancer progression correlates with increased Wnt5a expression in TAMs [77]. These observations, combined with studies implicating TEMs as the predominant proangiogenic macrophage within the tumor microenvironment [47,68], suggest that this model may be relevant to tumor angiogenesis as well, especially in light of the tumor being a “wound that does not heal” [91]. Owing to the perinatal lethality exhibited by Wnt5a-null mice [92], it has been difficult to study its function in vivo. Until conditional Wnt5a KO mice are made, we anticipate that complex three-dimensional in vitro models that incorporate macrophages and allow for easy manipulation of cells will be vital in elucidating the role that this signaling pathway plays in macrophage-regulation of angiogenesis. Looking ahead, we foresee more sophisticated experimental systems uncovering an even greater role for macrophage-derived Wnt ligands in regulating angiogenesis.

### Competing interests

The authors declare no competing interests.

### Authors' contributions

ACN and CCWH wrote the paper. Both authors read and approved the final manuscript.

### Acknowledgements

NIH RO1 HL60067  
Edwards Lifesciences Center for Advanced Cardiovascular Technology  
Graduate Student Fellowship  
Chao Family Comprehensive Cancer Center

### Author details

<sup>1</sup>The Department of Molecular Biology and Biochemistry, University of California Irvine, Irvine, CA 92697, USA. <sup>2</sup>The Department of Biomedical Engineering, University of California Irvine, Irvine, CA 92697, USA. <sup>3</sup>The Edwards Lifesciences Center for Advanced Cardiovascular Technology, University of California Irvine, Irvine, CA 92697, USA.

Received: 23 June 2012 Accepted: 13 July 2012

Published: 31 August 2012

### References

1. Carmeliet P, Jain RK: Molecular mechanisms and clinical applications of angiogenesis. *Nature* 2011, **473**:298–307.
2. Hughes CC: Endothelial-stromal interactions in angiogenesis. *Curr Opin Hematol* 2008, **15**:204–209.
3. Clevers H: Wnt/beta-catenin signaling in development and disease. *Cell* 2006, **127**:469–480.
4. Pollard JW: Trophic macrophages in development and disease. *Nat Rev Immunol* 2009, **9**:259–270.
5. Gordon S, Martinez FO: Alternative activation of macrophages: mechanism and functions. *Immunity* 2010, **32**:593–604.
6. Sica A, Mantovani A: Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest* 2012, **122**:787–795.

7. Angers S, Moon RT: Proximal events in Wnt signal transduction. *Nat Rev Mol Cell Biol* 2009, **10**:468–477.
8. van Amerongen R, Nusse R: Towards an integrated view of Wnt signaling in development. *Development* 2009, **136**:3205–3214.
9. Mikels AJ, Nusse R: Purified Wnt5a protein activates or inhibits beta-catenin-TCF signaling depending on receptor context. *PLoS Biol* 2006, **4**:e115.
10. Miller JR: The Wnts. *Genome Biol* 2002, **3**: REVIEWS3001.
11. van Amerongen R, Mikels A, Nusse R: Alternative wnt signaling is initiated by distinct receptors. *Sci Signal* 2008, **1**:re9.
12. Seifert JR, Mlodzik M: Frizzled/PCP signalling: a conserved mechanism regulating cell polarity and directed motility. *Nat Rev Genet* 2007, **8**:126–138.
13. Goodwin AM, Sullivan KM, D'Amore PA: Cultured endothelial cells display endogenous activation of the canonical Wnt signaling pathway and express multiple ligands, receptors, and secreted modulators of Wnt signaling. *Dev Dyn* 2006, **235**:3110–3120.
14. Favre CJ, Mancuso M, Maas K, McLean JW, Baluk P, McDonald DM: Expression of genes involved in vascular development and angiogenesis in endothelial cells of adult lung. *Am J Physiol Heart Circ Physiol* 2003, **285**:H1917–H1938.
15. van Gijn ME, Blankesteijn WM, Smits JF, Hierck B, Gittenberger-de Groot AC: Frizzled 2 is transiently expressed in neural crest-containing areas during development of the heart and great arteries in the mouse. *Anat Embryol (Berl)* 2001, **203**:185–192.
16. Mao C, Malek OT, Pueyo ME, Steg PG, Soubrier F: Differential expression of rat frizzled-related frzb-1 and frizzled receptor fz1 and fz2 genes in the rat aorta after balloon injury. *Arterioscler Thromb Vasc Biol* 2000, **20**:43–51.
17. Wright M, Aikawa M, Szeto W, Papkoff J: Identification of a Wnt-responsive signal transduction pathway in primary endothelial cells. *Biochem Biophys Res Commun* 1999, **263**:384–388.
18. Masckauchan TN, Shawber CJ, Funahashi Y, Li CM, Kitajewski J: Wnt/beta-catenin signaling induces proliferation, survival and interleukin-8 in human endothelial cells. *Angiogenesis* 2005, **8**:43–51.
19. Masckauchan TN, Agalliu D, Vorontchikhina M, Ahn A, Parmalee NL, Li CM, Khoo A, Tycko B, Brown AM, Kitajewski J: Wnt5a signaling induces proliferation and survival of endothelial cells in vitro and expression of MMP-1 and Tie-2. *Mol Biol Cell* 2006, **17**:5163–5172.
20. Goodwin AM, Kitajewski J, D'Amore PA: Wnt1 and Wnt5a affect endothelial proliferation and capillary length; Wnt2 does not. *Growth Factors* 2007, **25**:25–32.
21. de Jesus Perez VA, Alastalo TP, Wu JC, Axelrod JD, Cooke JP, Amieva M, Rabinovitch M: Bone morphogenetic protein 2 induces pulmonary angiogenesis via Wnt-beta-catenin and Wnt-RhoA-Rac1 pathways. *J Cell Biol* 2009, **184**:83–99.
22. Cheng CW, Yeh JC, Fan TP, Smith SK, Charnock-Jones DS: Wnt5a-mediated non-canonical Wnt signalling regulates human endothelial cell proliferation and migration. *Biochem Biophys Res Commun* 2008, **365**:285–290.
23. Cheng CW, Smith SK, Charnock-Jones DS: Wnt-1 signaling inhibits human umbilical vein endothelial cell proliferation and alters cell morphology. *Exp Cell Res* 2003, **291**:415–425.
24. Ezan J, Leroux L, Barandon L, Dufourcq P, Jaspard B, Moreau C, Allieres C, Daret D, Couffignal T, Duplaa C: FrzA/sFRP-1, a secreted antagonist of the Wnt-Frizzled pathway, controls vascular cell proliferation in vitro and in vivo. *Cardiovasc Res* 2004, **63**:731–738.
25. Wang H, Charles PC, Wu Y, Ren R, Pi X, Moser M, Barshishat-Kupper M, Rubin JS, Perou A, Bautch V, Patterson C: Gene expression profile signatures indicate a role for Wnt signaling in endothelial commitment from embryonic stem cells. *Circ Res* 2006, **98**:1331–1339.
26. Blankesteijn WM, van Gijn ME, Essers-Janssen YP, Daemen MJ, Smits JF: Beta-catenin, an inducer of uncontrolled cell proliferation and migration in malignancies, is localized in the cytoplasm of vascular endothelium during neovascularization after myocardial infarction. *Am J Pathol* 2000, **157**:877–883.
27. Daneman R, Agalliu D, Zhou L, Kuhnert F, Kuo CJ, Barres BA: Wnt/beta-catenin signaling is required for CNS, but not non-CNS, angiogenesis. *Proc Natl Acad Sci U S A* 2009, **106**:641–646.
28. Eberhart CG, Argani P: Wnt signaling in human development: beta-catenin nuclear translocation in fetal lung, kidney, placenta, capillaries, adrenal, and cartilage. *Pediatr Dev Pathol* 2001, **4**:351–357.
29. Eberhart CG, Tihan T, Burger PC: Nuclear localization and mutation of beta-catenin in medulloblastomas. *J Neuropathol Exp Neurol* 2000, **59**:333–337.
30. Maretto S, Cordenonsi M, Dupont S, Braghetta P, Broccoli V, Hassan AB, Volpin D, Bressan GM, Piccolo S: Mapping Wnt/beta-catenin signaling during mouse development and in colorectal tumors. *Proc Natl Acad Sci U S A* 2003, **100**:3299–3304.
31. Phng LK, Potente M, Leslie JD, Babbage J, Nyqvist D, Lobov I, Ondr JK, Rao S, Lang RA, Thurston G, Gerhardt H: Nrarp coordinates endothelial Notch and Wnt signaling to control vessel density in angiogenesis. *Dev Cell* 2009, **16**:70–82.
32. Yano H, Hara A, Shinoda J, Takenaka K, Yoshimi N, Mori H, Sakai N: Immunohistochemical analysis of beta-catenin in N-ethyl-N-nitrosourea-induced rat gliomas: implications in regulation of angiogenesis. *Neurol Res* 2000, **22**:527–532.
33. Yano H, Hara A, Takenaka K, Nakatani K, Shinoda J, Shimokawa K, Yoshimi N, Mori H, Sakai N: Differential expression of beta-catenin in human glioblastoma multiforme and normal brain tissue. *Neurol Res* 2000, **22**:650–656.
34. Corada M, Nyqvist D, Orsenigo F, Caprini A, Giampietro C, Taketo MM, Iruela-Arispe ML, Adams RH, Dejana E: The Wnt/beta-catenin pathway modulates vascular remodeling and specification by upregulating Dll4/Notch signaling. *Dev Cell* 2010, **18**:938–949.
35. Nimmagadda S, Geetha-Loganathan P, Scaal M, Christ B, Huang R: FGFs, Wnts and BMPs mediate induction of VEGFR-2 (Quek-1) expression during avian somite development. *Dev Biol* 2007, **305**:421–429.
36. Monkley SJ, Delaney SJ, Pennisi DJ, Christiansen JH, Wainwright BJ: Targeted disruption of the Wnt2 gene results in placental defects. *Development* 1996, **122**:3343–3353.
37. Robitaille J, MacDonald ML, Kaykas A, Sheldahl LC, Zeisler J, Dube MP, Zhang LH, Singaraja RR, Guernsey DL, Zheng B, et al: Mutant frizzled-4 disrupts retinal angiogenesis in familial exudative vitreoretinopathy. *Nat Genet* 2002, **32**:326–330.
38. Ye X, Wang Y, Cahill H, Yu M, Badea TC, Smallwood PM, Peachey NS, Nathans J: Norrin, frizzled-4, and Lrp5 signaling in endothelial cells controls a genetic program for retinal vascularization. *Cell* 2009, **139**:285–298.
39. Descamps B, Sewduth R, Ferreira Tojais N, Jaspard B, Reynaud A, Sohet F, Lacolley P, Allieres C, Lamaziere JM, Moreau C, et al: Frizzled 4 regulates arterial network organization through noncanonical Wnt/planar cell polarity signaling. *Circ Res* 2012, **110**:47–58.
40. Ishikawa T, Tamai Y, Zorn AM, Yoshida H, Seldin MF, Nishikawa S, Taketo MM: Mouse Wnt receptor gene Fzd5 is essential for yolk sac and placental angiogenesis. *Development* 2001, **128**:25–33.
41. Liu C, Nathans J: An essential role for frizzled 5 in mammalian ocular development. *Development* 2008, **135**:3567–3576.
42. Dejana E: The role of wnt signaling in physiological and pathological angiogenesis. *Circ Res* 2010, **107**:943–952.
43. Franco CA, Liebner S, Gerhardt H: Vascular morphogenesis: a Wnt for every vessel? *Curr Opin Genet Dev* 2009, **19**:476–483.
44. Tsaousi A, Mill C, George SJ: The Wnt pathways in vascular disease: lessons from vascular development. *Curr Opin Lipidol* 2011, **22**:350–357.
45. Nucera S, Biziato D, De Palma M: The interplay between macrophages and angiogenesis in development, tissue injury and regeneration. *Int J Dev Biol* 2011, **55**:495–503.
46. Qian BZ, Pollard JW: Macrophage diversity enhances tumor progression and metastasis. *Cell* 2010, **141**:39–51.
47. Pucci F, Venneri MA, Biziato D, Nonis A, Moi D, Sica A, Di Serio C, Naldini L, De Palma M: A distinguishing gene signature shared by tumor-infiltrating Tie2-expressing monocytes, blood "resident" monocytes, and embryonic macrophages suggests common functions and developmental relationships. *Blood* 2009, **114**:901–914.
48. Rae F, Woods K, Sasmono T, Campanale N, Taylor D, Ovchinnikov DA, Grimmond SM, Hume DA, Ricardo SD, Little MH: Characterisation and trophic functions of murine embryonic macrophages based upon the use of a Csf1r-EGFP transgene reporter. *Dev Biol* 2007, **308**:232–246.
49. Fantin A, Vieira JM, Gestri G, Denti L, Schwarz Q, Prykhodzhiy S, Peri F, Wilson SW, Ruhrberg C: Tissue macrophages act as cellular chaperones for vascular anastomosis downstream of VEGF-mediated endothelial tip cell induction. *Blood* 2010, **116**:829–840.
50. Outtz HH, Tattersall IW, Kofler NM, Steinbach N, Kitajewski J: Notch1 controls macrophage recruitment and Notch signaling is activated at sites of endothelial cell anastomosis during retinal angiogenesis in mice. *Blood* 2011, **118**:3436–3439.

51. Rymo SF, Gerhardt H, Wolfhagen Sand F, Lang R, Uv A, Betsholtz C: **A two-way communication between microglial cells and angiogenic sprouts regulates angiogenesis in aortic ring cultures.** *PLoS One* 2011, **6**:e15846.
52. Moldovan NI, Goldschmidt-Clermont PJ, Parker-Thornburg J, Shapiro SD, Kolattukudy PE: **Contribution of monocytes/macrophages to compensatory neovascularization: the drilling of metalloelastase-positive tunnels in ischemic myocardium.** *Circ Res* 2000, **87**:378–384.
53. Lucas T, Waisman A, Ranjan R, Roes J, Krieg T, Muller W, Roers A, Eming SA: **Differential roles of macrophages in diverse phases of skin repair.** *J Immunol* 2010, **184**:3964–3977.
54. Okuno Y, Nakamura-Ishizu A, Kishi K, Suda T, Kubota Y: **Bone marrow-derived cells serve as proangiogenic macrophages but not endothelial cells in wound healing.** *Nature* 2011, **475**:222–225.
55. Mantovani A, Sica A: **Macrophages, innate immunity and cancer: balance, tolerance, and diversity.** *Curr Opin Immunol* 2010, **22**:231–237.
56. Zumsteg A, Christofori G: **Corrupt policemen: inflammatory cells promote tumor angiogenesis.** *Curr Opin Oncol* 2009, **21**:60–70.
57. Qian BZ, Li J, Zhang H, Kitamura T, Zhang J, Campion LR, Kaiser EA, Snyder LA, Pollard JW: **CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis.** *Nature* 2011, **475**:222–225.
58. Scholl SM, Pallud C, Beuvon F, Hacene K, Stanley ER, Rohrschneider L, Tang R, Pouillart P, Lidereau R: **Anti-colony-stimulating factor-1 antibody staining in primary breast adenocarcinomas correlates with marked inflammatory cell infiltrates and prognosis.** *J Natl Cancer Inst* 1994, **86**:120–126.
59. Kubota Y, Takubo K, Shimizu T, Ohno H, Kishi K, Shibuya M, Saya H, Suda T: **M-CSF inhibition selectively targets pathological angiogenesis and lymphangiogenesis.** *J Exp Med* 2009, **206**:1089–1102.
60. Lin EY, Li JF, Gnatovskiy L, Deng Y, Zhu L, Grzesik DA, Qian H, Xue XN, Pollard JW: **Macrophages regulate the angiogenic switch in a mouse model of breast cancer.** *Cancer Res* 2006, **66**:11238–11246.
61. Lin EY, Pollard JW: **Tumor-associated macrophages press the angiogenic switch in breast cancer.** *Cancer Res* 2007, **67**:5064–5066.
62. Jin H, Su J, Garmy-Susini B, Kleeman J, Varner J: **Integrin alpha4beta1 promotes monocyte trafficking and angiogenesis in tumors.** *Cancer Res* 2006, **66**:2146–2152.
63. Leek RD, Harris AL: **Tumor-associated macrophages in breast cancer.** *J Mammary Gland Biol Neoplasia* 2002, **7**:177–189.
64. Giraudo E, Inoue M, Hanahan D: **An amino-bisphosphonate targets MMP-9-expressing macrophages and angiogenesis to impair cervical carcinogenesis.** *J Clin Invest* 2004, **114**:623–633.
65. Nocito A, Dahm F, Jochum W, Jang JH, Georgiev P, Bader M, Graf R, Clavien PA: **Serotonin regulates macrophage-mediated angiogenesis in a mouse model of colon cancer allografts.** *Cancer Res* 2008, **68**:5152–5158.
66. Du R, Lu KV, Petritsch C, Liu P, Ganss R, Passegue E, Song H, Vandenberg S, Johnson RS, Werb Z, Bergers G: **HIF1alpha induces the recruitment of bone marrow-derived vascular modulatory cells to regulate tumor angiogenesis and invasion.** *Cancer Cell* 2008, **13**:206–220.
67. Stockmann C, Doedens A, Weidemann A, Zhang N, Takeda N, Greenberg JI, Chersesh DA, Johnson RS: **Deletion of vascular endothelial growth factor in myeloid cells accelerates tumorigenesis.** *Nature* 2008, **456**:814–818.
68. De Palma M, Venneri MA, Galli R, Sergi L, Politi LS, Sampaoli M, Naldini L: **Tie2 identifies a hematopoietic lineage of proangiogenic monocytes required for tumor vessel formation and a mesenchymal population of pericyte progenitors.** *Cancer Cell* 2005, **8**:211–226.
69. Coffelt SB, Tal AO, Scholz A, De Palma M, Patel S, Urbich C, Biswas SK, Murdoch C, Plate KH, Reiss Y, Lewis CE: **Angiopoietin-2 regulates gene expression in Tie2-expressing monocytes and augments their inherent proangiogenic functions.** *Cancer Res* 2010, **70**:5270–5280.
70. Murdoch C, Tazzyman S, Webster S, Lewis CE: **Expression of Tie-2 by human monocytes and their responses to angiopoietin-2.** *J Immunol* 2007, **178**:7405–7411.
71. Pereira C, Schaer DJ, Bachli EB, Kurrer MO, Schoedon G: **Wnt5A/CaMKII signaling contributes to the inflammatory response of macrophages and is a target for the antiinflammatory action of activated protein C and interleukin-10.** *Arterioscler Thromb Vasc Biol* 2008, **28**:504–510.
72. Campbell IL, Abraham CR, Masliah E, Kemper P, Inglis JD, Oldstone MB, Mucke L: **Neurologic disease induced in transgenic mice by cerebral overexpression of interleukin 6.** *Proc Natl Acad Sci U S A* 1993, **90**:10061–10065.
73. Fan Y, Ye J, Shen F, Zhu Y, Yeghiazarians Y, Zhu W, Chen Y, Lawton MT, Young WL, Yang GY: **Interleukin-6 stimulates circulating blood-derived endothelial progenitor cell angiogenesis in vitro.** *J Cereb Blood Flow Metab* 2008, **28**:90–98.
74. Li A, Dubey S, Varney ML, Dave BJ, Singh RK: **IL-8 directly enhanced endothelial cell survival, proliferation, and matrix metalloproteinases production and regulated angiogenesis.** *J Immunol* 2003, **170**:3369–3376.
75. Rosell A, Arai K, Lok J, He T, Guo S, Navarro M, Montaner J, Katusic ZS, Lo EH: **Interleukin-1beta augments angiogenic responses of murine endothelial progenitor cells in vitro.** *J Cereb Blood Flow Metab* 2009, **29**:933–943.
76. Kim J, Kim DW, Ha Y, Ihm MH, Kim H, Song K, Lee I: **Wnt5a induces endothelial inflammation via beta-catenin-independent signaling.** *J Immunol* 2010, **185**:1274–1282.
77. Smith K, Bui TD, Poulosom R, Kaklamanis L, Williams G, Harris AL: **Up-regulation of macrophage wnt gene expression in adenoma-carcinoma progression of human colorectal cancer.** *Br J Cancer* 1999, **81**:496–502.
78. Staton CA, Chetwood AS, Cameron IC, Cross SS, Brown NJ, Reed MW: **The angiogenic switch occurs at the adenoma stage of the adenoma carcinoma sequence in colorectal cancer.** *Gut* 2007, **56**:1426–1432.
79. Wyckoff JB, Segall JE, Condeelis JS: **The collection of the motile population of cells from a living tumor.** *Cancer Res* 2000, **60**:5401–5404.
80. Ojalvo LS, Whittaker CA, Condeelis JS, Pollard JW: **Gene expression analysis of macrophages that facilitate tumor invasion supports a role for Wnt-signaling in mediating their activity in primary mammary tumors.** *J Immunol* 2010, **184**:702–712.
81. Gergely K, Gerinac A: **A consonant construction of the hyaloid and retinal vascular systems by the angiogenic process.** *Bratisl Lek Listy* 2011, **112**:143–151.
82. Diez-Roux G, Argilla M, Makarenkova H, Ko K, Lang RA: **Macrophages kill capillary cells in G1 phase of the cell cycle during programmed vascular regression.** *Development* 1999, **126**:2141–2147.
83. Lobov IB, Rao S, Carroll TJ, Vallance JE, Ito M, Ondr JK, Kurup S, Glass DA, Patel MS, Shu W, et al: **WNT7b mediates macrophage-induced programmed cell death in patterning of the vasculature.** *Nature* 2005, **437**:417–421.
84. Rao S, Lobov IB, Vallance JE, Tsujikawa K, Shiojima I, Akunuru S, Walsh K, Benjamin LE, Lang RA: **Obligatory participation of macrophages in an angiopoietin 2-mediated cell death switch.** *Development* 2007, **134**:4449–4458.
85. Saint-Geniez M, D'Amore PA: **Development and pathology of the hyaloid, choroidal and retinal vasculature.** *Int J Dev Biol* 2004, **48**:1045–1058.
86. Stefater JA 3rd, Lewkowich I, Rao S, Mariggi G, Carpenter AC, Burr AR, Fan J, Ajima R, Molkentin JD, Williams BO, et al: **Regulation of angiogenesis by a non-canonical Wnt-Flt1 pathway in myeloid cells.** *Nature* 2011, **474**:511–515.
87. Ching W, Nusse R: **A dedicated Wnt secretion factor.** *Cell* 2006, **125**:432–433.
88. Kendall RL, Thomas KA: **Inhibition of vascular endothelial cell growth factor activity by an endogenously encoded soluble receptor.** *Proc Natl Acad Sci U S A* 1993, **90**:10705–10709.
89. Shibuya M: **Structure and dual function of vascular endothelial growth factor receptor-1 (Flt-1).** *Int J Biochem Cell Biol* 2001, **33**:409–420.
90. Crampton SP, Wu B, Park EJ, Kim JH, Solomon C, Waterman ML, Hughes CC: **Integration of the beta-catenin-dependent Wnt pathway with integrin signaling through the adaptor molecule Grb2.** *PLoS One* 2009, **4**:e7841.
91. Dvorak HF: **Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing.** *N Engl J Med* 1986, **315**:1650–1659.
92. Yamaguchi TP, Bradley A, McMahon AP, Jones S: **A Wnt5a pathway underlies outgrowth of multiple structures in the vertebrate embryo.** *Development* 1999, **126**:1211–1223.

doi:10.1186/2045-824X-4-13

Cite this article as: Newman and Hughes: Macrophages and angiogenesis: a role for Wnt signaling. *Vascular Cell* 2012 **4**:13.